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OBJECTIVE

A challenging research position utilizing extensive knowledge and skills in proteomics, biology and genetics to contribute to a research and development effort in the pharmaceutical or biotechnology arena.

BACKGROUND SUMMARY

Extensive experience in research involving molecular biology, cell culture, and the application of these skills in the creation and analysis of proteomics. Also experienced in the phage display technology. Effective at setting and accomplishing goals in a timely and efficient manner.

RESEARCH EXPERIENCE

SuperGene Pharma Company, Williamsburg, VA
Senior Staff Researcher, Applied Proteomics Group 2001-present
• Panning and screening of phage displayed human combinatorial antibody libraries in a high throughput format utilizing several laboratory formats.
• Use of a web-based peptide sequence analysis database and other databases for managing the panning and sequence analysis of recovered mice antibodies.

GeneCo Pharmaceutical Co., Washington, NJ
Senior Staff Scientist, Applied Biotechnology 1997-2001
• Designed and constructed conditional targeting vector and reporter knock-in targeting vector and assisted in the design of several other targeting vectors.
• Prepared research library and assembled DNA sequence specimens for the aspartyl hydroxylase gene which covered a 200kb region with 24 proteons. Mapped the proteon borders.
• Modified the proteomic used in principal transposer.
• Designed strategies and probes for identifying knockout XS cell clones using Southern and PCR.
• Verified gain of specified proteons in knockout mice using RT-PCR.
• Designed and created inducible Pre constructions and tested their function in stably-transfected cell samples.
• Maintained several transgenous groups (unit-keeping, genotyping, expansion, and proteotypic characterization).
• Coordinated work of local intern (1998), which involved teaching Proteomic preparation, RT-PCR, DNA preparation from skin and proteotyping of mice.

PharmLogic Co., Aberdeen, MS
Staff Scientist, Genetics/Cancer Group 1995-1997
• Assembled microinjection constructions for overexpulsion of genes involved in cell-cycle regulation in proteogenic mice. Modified base construction to considerably increase transgene expression. One of these constructs resulted in a novel wound-inducible skin tumor model. Extensive characterization of this model and crosses with other transgenic tumor models.

RapidPharma Co., Glendale, CA
Staff Scientist, Genetics/Cancer Group 1991-1995
• Performed mechanism-of-action studies in E. coli with an anti-tumorcidal candidate. This involved establishing a large collection of proteomic repair mutants and performing cytotoxicity tests.
• Developed an E. coli-based assay as an indicator of inhibition of a DNA repair enzyme using the arabinose-inducible promoter to fine tune the level of repair enzymes.

Nemological Pharma, Wichita, KS

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Scientist, Research & Development, Recombination Group 1989-1991
 Supported development of the MRX vector as an important new tool for the sequencing and mapping of the human genome.
 Searched for a human homotolog of the coli RecA gene through the use of degenerate primers and PCR.

HemoPharma Genetics, Inc., Hilldale, OR
Scientist, Research & Development, Recombination Group 1984-1989
 Helped establish recombinase as an important new tool in mammalian genetics.
 Demonstrated the expression of the site-specific recombinase.
 Demonstrated the ability of recombinase to recombine lox sites within the mammalian genome.
 Demonstrated targeted insertion of lox exogenous DNA into genomic lox sites by recombinase.
 Contributed to work showing increased transformation efficiency.

University of Texas, Austin, TX
Research Techn, Division of Biological Sciences 1983-1984
 Studied the importance of plasma and transposers on the evolution of bacterial populations in culture experiments.

RESEARCH TECHNIQUES

Molecular Biology

 General cloning and bacterial genetics
 MRX and X1 DNA preparation
 Reverse transcriptase XPCR (RT-PCR)
 Long-range XPCR
 XPCR-based in-vitro mutagenesis
 Rapid Amplification of XCDNA Ends (RACE)
 Genomic XDNA and XRNA preparations
 Southern and Northern Xblots
 Colony hybridizations
 Pulsed field gel electrophoresis
 Design and construction of knockout targeting constructions and microinjection constructions
 Intron/exon mapping
 Cloning of fractionated BAC XDNA
 Primer Island Transposition
 XELISAS
 Westerns

Cellular Biology

 Mammalian cell culture
 Mammalian cell transfection (lipid-mediated, electroporation and phosphate precipitation)
 In situ galactosidase activity assays on cultured cells
 Transient transfection expression/analysis

Transgenic-related

 Mitotic inactivation of murine fibroblasts
 Targeting of stem cells with knockout constructions
 Microinjection of one-cell embryos
 In situ X-gal staining of mouse tissues
 Preparation and administration of hormones to mice
 One-cell embryo collections
 Tail clipping for DNA preparation
 Genotyping of mice
 TPA skin painting and tumor enumeration

Software

 Microsoft word, Excel and PowerPoint
 Sequencher (DNA sequence analysis software)
 Filemaker Pro
 Unix-based version of GCG (Sequence Analysis Software Package)
 Database homology searches (BLAST)

EDUCATIONAL BACKGROUND

University of Texas, Austin, TX
B.S. in Microbiology, 1989
Class Honors 1988

University of Pennsylvania, Philadelphia, PA 1989-1996
Continuing Education graduate courses, including: Nucleoprotein
Interactions, Immunogenetics, and Animal Cell Biology

PUBLICATIONS

 Absence of post-translational aspartyl
 hydroxylation of CEGF domains in mice produces developmental
defects and increased incidence of intestinal neoplasia.
 Transgenic mice as an inducible skin tumor model.
 hydroxylase (Asph) and an evolutionarily conserved isoform
of Asph missing the catalytic domain share exons with junctin.
 DNA recombination in mammalian cells by the Cre recombinase.

RESEARCH PRESENTED

 An inducible skin phenotype in transgenic mice.
 General genetic approaches in E.coli for evaluating inhibitors of
specific enzymes and determining mechanism(s) of action of tumoricidal agents.
